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# How structural elements evolving from bacterial to human SLC6 transporters enabled new functional properties

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## Abstract

**Background:** Much of the structure-based mechanistic understandings of the function of SLC6A neurotransmitter transporters emerged from the study of their bacterial LeuT-fold homologs. It has become evident, however, that structural differences such as the long N- and C-termini of the eukaryotic neurotransmitter transporters are involved in an expanded set of functional properties to the eukaryotic transporters. These functional properties are not shared by the bacterial homologs, which lack the structural elements that appeared later in evolution. However, mechanistic insights into some of the measured functional properties of the eukaryotic transporters that have been suggested to involve these structural elements are sparse or merely descriptive.

**Results:** To learn how the structural elements added in evolution enable mechanisms of the eukaryotic transporters in ways not shared with their bacterial LeuT-like homologs, we focused on the human dopamine transporter (hDAT) as a prototype. We present the results of a study employing large-scale molecular dynamics simulations and comparative Markov state model analysis of experimentally determined properties of the wild-type and mutant hDAT constructs. These offer a quantitative outline of mechanisms in which a rich spectrum of interactions of the hDAT N-terminus and C-terminus contribute to the regulation of transporter function (e.g., by phosphorylation) and/or to entirely new phenotypes (e.g., reverse uptake (efflux)) that were added in evolution.

**Conclusions:** The findings are consistent with the proposal that the size of eukaryotic neurotransmitter transporter termini increased during evolution to enable more functions (e.g., efflux) not shared with the bacterial homologs. The mechanistic explanations for the experimental findings about the modulation of function in DAT, the serotonin transporter, and other eukaryotic transporters reveal separate roles for the distal and proximal segments of the much larger N-terminus in eukaryotic transporters compared to the bacterial ones. The involvement of the *proximal* and *distal* segments — such as the role of the *proximal* segment in sustaining transport in phosphatidylinositol 4,5-bisphosphate-depleted membranes and of the *distal* segment in modulating efflux — may represent an evolutionary adaptation required for the function of eukaryotic transporters expressed in various cell types of the same organism that differ in the lipid composition and protein complement of their membrane environment.

**Keywords:** Dopamine transport, SLC6 neurotransmitter transporters, Evolutionary gain of function, Molecular dynamics simulations, Markov state models, Reverse transport, SERT, Regulation by PIP<sub>2</sub>, Regulation by phosphorylation, posttranslational modifications

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## Background

The dopamine transporter (DAT) is a member of the neurotransmitter:sodium symporter (NSS) family of proteins belonging to the solute carrier 6 (SLC6) family that performs the reuptake of dopamine from the synaptic cleft into the presynaptic nerve required for neuronal signaling [1]. The essential role of DAT, and of its closely related homologs — the serotonin and norepinephrine transporters (SERT and NET) — in signal termination at the synapse makes them important targets for psychostimulants such as cocaine and amphetamines, as well as for pharmaceutical treatment of a variety of disorders of the nervous system [2]. Moreover, genetic modifications of the functions of these transporters [3, 4] have been implicated in diseases including schizophrenia, Parkinson's disease, and attention-deficit/hyperactivity disorder (ADHD). Reverse transport of the neurotransmitters (efflux) mediated by DAT and SERT, which has been shown to be affected by such disease-related mutations, is currently a very active topic of research on mechanisms of these membrane proteins [4–15].

Much has been learned about these mammalian neurotransmitter transporters from the investigation of the structure and function of their bacterial homologs, with which they share many structural and mechanistic properties [16–19]. It has become clear, however, that important structural differences exist between the eukaryotic and bacterial proteins, the largest being the much longer N- and C-termini that have been proposed to be partially structured [20, 21]. Notably, experimental data point to an involvement of these regions of structural difference in measured functional properties of the mammalian NSS [22–24]. For example, the phosphorylation of the N-terminus has been implicated in the efflux functions of the human DAT (hDAT) [22, 25], and our work has shown that the amphetamine (AMPH)-induced reverse transport (efflux) exhibited by DAT and SERT, but not by the bacterial analogs, is dependent on electrostatic interactions between the hDAT N-terminus and negatively charged phosphatidylinositol 4,5-bisphosphate (PIP<sub>2</sub>) lipids in the membrane [34, 41]. Notably, the measured substrate uptake by DAT, a function it shares with the bacterial transporters, is not affected by this N-terminus interaction [34].

Structure-based mechanistic details of the mode in which eukaryotic NSS function is modulated by the involvement of the N-terminus are still sparse, but mechanistic insight from computational studies of hDAT has shown [29] that the N-terminus of DAT engages the PIP<sub>2</sub> lipid component of membranes to achieve *conformational changes related to function* (CCRF). These include (1) the transition of the DAT from outward-facing to inward-facing configurations [30] and (2) the release of the sodium ion from the Na<sup>2</sup> binding site that is known to precede substrate transport

[26, 31] and can serve as a monitor of the initial stages of the functional mechanism [32]. The complexity of the different CCRF suggested that the N-terminus/PIP<sub>2</sub> interactions supporting them may (1) involve different modes of association with the membrane under various conditions (such as in the presence/absence of PIP<sub>2</sub>) and (2) would be different for different hDAT constructs (e.g., when the N-terminus is phosphorylated or mutated). This reasoning provided a specific testable hypothesis about the mechanism by which new properties of the eukaryotic transporters, those that are not shared with the bacterial homologs, are enabled by the addition of the long N- and C-terminals.

To probe this hypothesis and verify the relation between the experimental measurements and the specific modes in which the N-terminus participates, we undertook the present computational study of the modes of interaction of the N-terminus in hDAT with the rest of the structure, including the C-terminus, under various conditions and with modifications (phosphorylation, mutations) that have known functional consequences. Here we describe the results from extensive ensemble-level all-atom molecular dynamics simulations we used in this study that also included Markov state model (MSM) analysis of hDAT dynamics modulated by mutated and/or modified N-terminus constructs and conditions. The results reveal preferred modes of interaction of the N-terminus with the intracellular domains of hDAT, which can be directly associated with experimentally measured functional phenotypes of the transporter. We show how these interaction patterns change under conditions that have been demonstrated to selectively affect efflux but not regular transport, e.g., PIP<sub>2</sub> depletion, mutations such as R51W, the K3A/K5A double mutation, or phosphomimic substitution, S/D, of serine residues at positions 2, 4, 7, 12, and 13 to aspartate [22, 33, 34]. Moreover, we verify, for the first time, the consistency of observations relating quantitative measures of the specific modes of interaction of the N-terminus with the measured functional properties attributed to them.

The coherent and direct relation between experimentally determined effects of the mutations and conditions and the interaction modes identified from the simulations validates the computational results and mechanistic conclusions. Moreover, because the mechanistic inferences are described in atomistic detail, they offer specific experimentally testable predictions for further studies of SLC6 transporter function and of the structure-based relation between the function of bacterial and eukaryotic members of this family. In particular, the details of the rich spectrum of modes of interaction of the long N-terminus of hDAT that emerges from these studies reveal the different roles of the *distal* and *proximal* segments of the N-terminus in modulating specific functions of hDAT. As these are

segments of the much larger N-terminus shared by the eukaryotic transporters compared to the bacterial ones, the findings bring mechanistic support for our proposal that the size of this region increased during evolution so as to enable more, and different, modes of regulation that are not shared with the bacterial analogs. An example discussed in detail is the mechanistic explanation for experimentally determined differences in the effects on uptake vs efflux resulting from manipulation of the N-terminus by partial truncation, mutations, and/or elimination of PIP<sub>2</sub> interactions. This example further underscores the central role of this structural addition in the evolution from the bacterial LeuT-like members of this family.

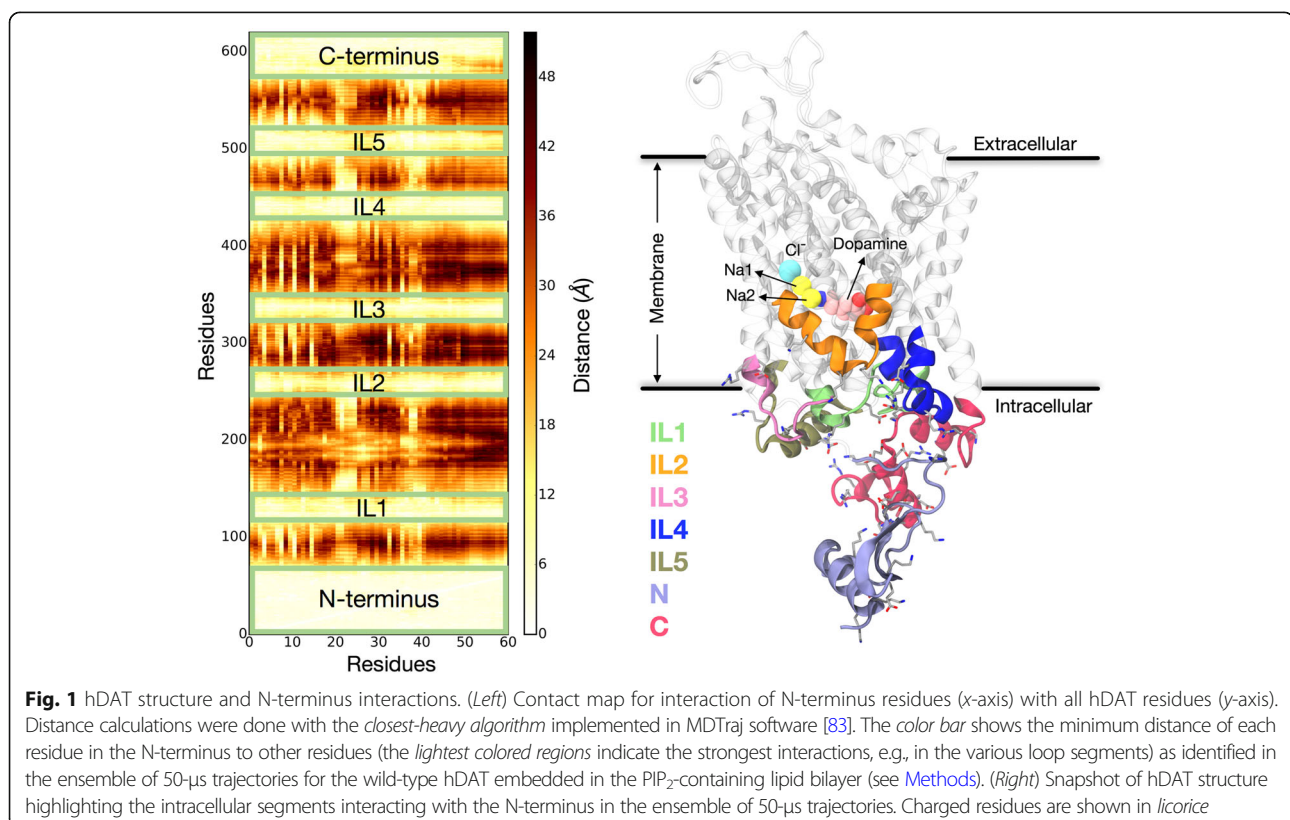
## Results

The full complement of molecular dynamics (MD) simulation trajectories carried out specifically for this study, as described in Methods, includes 50 statistically independent ~1- $\mu$ s-long trajectories for each of the four different conditions and constructs for which quantitative measurements of activity are available. As discussed before [32], the ensemble exploration of the configurational space of each construct/condition bolsters the statistical validity of the inferences and predictions from the simulation. The specific molecular systems investigated in this manner include (1) wild-type hDAT in PIP<sub>2</sub>-depleted

membranes (hereafter referred to as the “no-PIP<sub>2</sub> system”), (2) hDAT with the R51W mutation in the N-terminal domain, with the double K3A + K5A mutation in the N-terminus (termed the “K3/5A system”), and those with the first five N-terminal serine residues substituted by Asp as a phosphomimic (termed “S/D construct”); these three mutant constructs were immersed in PIP<sub>2</sub>-containing bilayers, consistent with the experimental conditions under which their functional properties were assayed. The data from these computations are compared to results for the wild-type hDAT simulated in PIP<sub>2</sub>-containing membranes we reported recently [32], and are analyzed utilizing the same protocols as described therein and detailed here in Methods.

### Different modes of interaction of the N-terminus with the rest of the hDAT protein correspond to differences in experimentally measured functional properties

From the simulation trajectories we identified the regions of hDAT structure that interact with the N-terminus, and generated the per-residue contact map shown in Fig. 1. This map shows that with PIP<sub>2</sub> present in the membrane, the N-terminus, as a whole, interacts with all intracellular loop (IL) regions of hDAT (IL1, IL2, IL3, IL4, and IL5), and with the C-terminus.

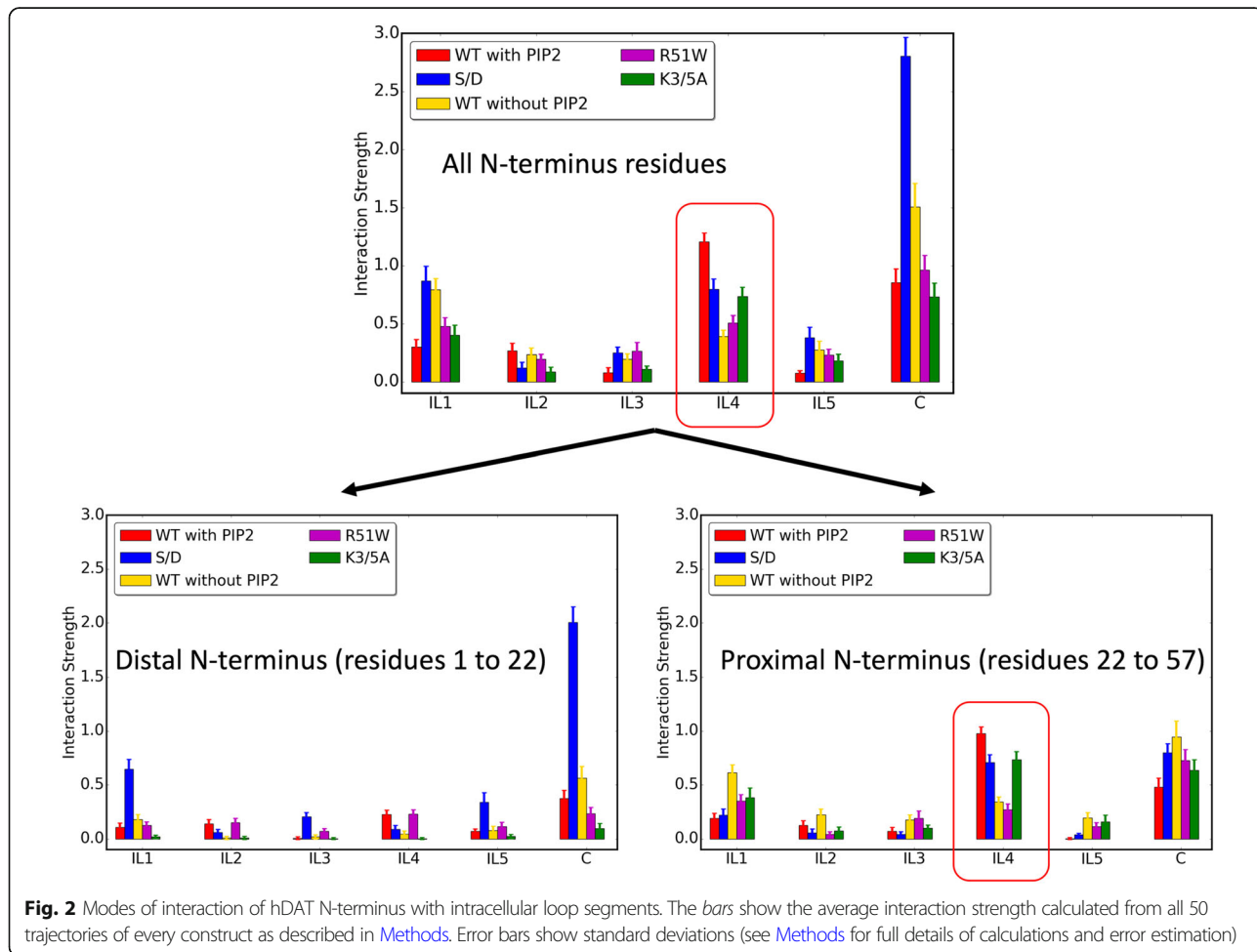


A detailed comparative analysis of the interactions between the N-terminus with the intracellular regions of wild-type hDAT in PIP<sub>2</sub>-containing membranes (obtained from equivalent trajectories described recently [32]) and the constructs studied here (including PIP<sub>2</sub>-depleted membrane conditions) reveals a specific pattern (modes) of interaction of the different parts of the N-terminus with intracellular regions of the transporter. These patterns are presented in Fig. 2.

As part of the experimental studies of reverse substrate transport (efflux) by DAT [22] and in SERT [35], their N-terminus was truncated, in the case of DAT eliminating the first 22 residues ( $\Delta$ N22 system). The measurements showed that the truncated transporters maintain direct substrate transport (uptake), but efflux is severely impaired. To compare with and interpret these experimental results, we calculated the modes of interaction of the various constructs for the corresponding components of the N-terminus: the *distal N-terminus* (residues 1–22) and the *proximal N-terminus* (residues 23–57). The results in Fig. 2 show a distinct difference between the interaction patterns of the two segments. Notably, the largest

differences are registered for the interactions of these *distal* and *proximal* segments with IL1, IL4, and the C-terminus. A remarkable similarity is revealed between the pattern of interactions with IL4 calculated for the entire N-terminus, and for just the *proximal* N-terminus (cf. the circled regions in Fig. 2). This similarity is especially noteworthy because the interaction with IL4 has been singled out to be essential in the early steps of the substrate transport mechanism marked by the release of Na<sup>+</sup> from the Na<sub>2</sub> site [29]. Thus, our finding here that the *proximal N-terminus* maintains the essential pattern of interaction with IL4 explains the surprising insensitivity of the inward substrate transport to the deletion of the first 22 residues.

The second largest difference between the interaction patterns of the *proximal* and *distal* N-terminal segments evident in Fig. 2 pertains to association with the C-terminus. In particular, the *distal* segment of the phosphomimic S/D construct has a remarkably high interaction quotient with the C-terminus, whereas the weakest association with the C-terminus is observed for the K3/5A construct (in fact, the distal segment of this double mutant K3A/K5A N-terminus is seen to engage in the least amount of interactions with



any of the intracellular loops). This diametrically opposed characteristic of the interaction of the two constructs relative to the wild type is remarkable and is fully consistent with the experimentally determined functional properties compared to the wild-type hDAT: (1) only the S/D mutant construct in which the *distal* N-terminus interacts strongly with the C-terminus has been found thus far to be able to enhance dopamine efflux mediated by DAT in the absence of AMPH [22], and (2) the K3/5A mutant, which we find to have the weakest interaction between the *distal* segment and the C-terminus, produces a very low AMPH-induced dopamine efflux [34]. The important relation of the *distal* segment with the efflux phenotype is underscored by the deleterious effect of the ( $\Delta$ N22) truncation on efflux, but not on uptake, as discussed above.

#### Multiple paths of inward release of $\text{Na}^+$ from the Na2 site are regulated by the modes of interaction of the N-terminus

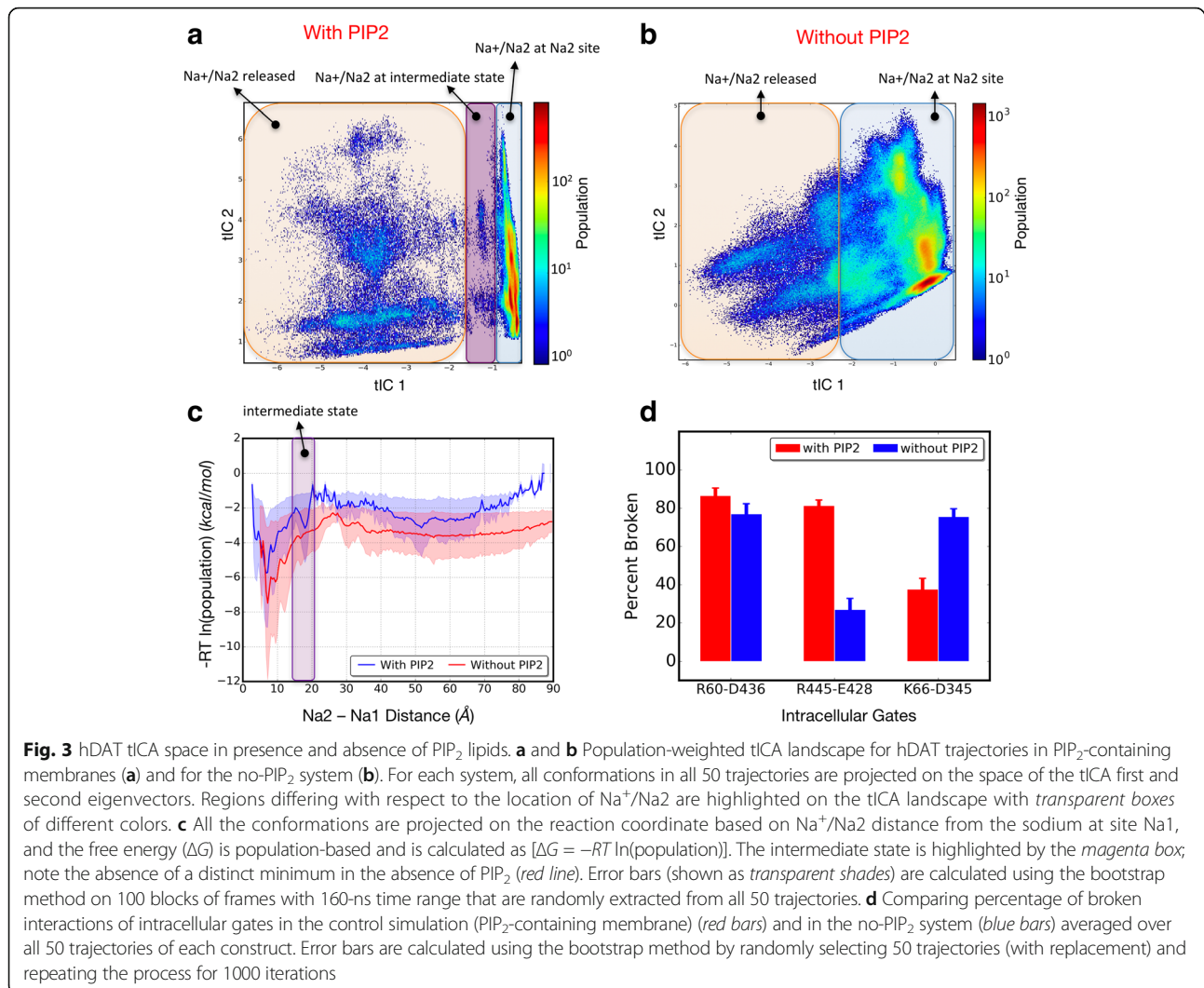
Our detailed study of the release of  $\text{Na}^+$  from the Na2 site (termed  $\text{Na}^+/\text{Na2}$  release) — which is known to initiate solute translocation by the hDAT [26, 31] — has identified CCRF and the underlying allosteric mechanism [27, 28, 32]. These CCRF relate directly to the mode of interaction of the N-terminus with intracellular regions of the transporter in  $\text{PIP}_2$ -containing membranes [32]. In the earlier studies [29, 32] we showed that specific  $\text{PIP}_2$ -mediated associations between the N-terminus and various intracellular loop regions of DAT trigger conformational transitions related to the release of  $\text{Na}^+/\text{Na2}$ . Here we find, from the new sets of MD simulations of the mutant constructs and conditions we study, that while they differ in their modes of N-terminus interactions (Fig. 2),  $\text{Na}^+/\text{Na2}$  release is observed, albeit at different rates, during simulations of the same time length for various systems (Additional file 1: Figures S1 to S4). While the rates of release events observed in the 50 trajectory ensembles for each construct/condition (see Additional file 1: Figures S1 to S4) are not rigorously comparable to each other in a statistically meaningful manner, the prediction of inward release of  $\text{Na}^+/\text{Na2}$  in each of these constructs is consistent with experimental results and with our previous finding [32] that the destabilization of  $\text{Na}^+/\text{Na2}$  is highly correlated with the amount of water penetration to the binding site (Additional file 2: Table S4). Additional file 1: Figures S5, S6, and S7 show details of the spontaneous release dynamics calculated for the S/D system (Additional file 1: Figure S5), the R51W hDAT system (Additional file 1: Figure S6), and the no- $\text{PIP}_2$  system (Additional file 1: Figure S7). The K3/5A mutant did not exhibit a release event, but the pattern of  $\text{Na}^+/\text{Na2}$  destabilization and intracellular gate opening is similar to that observed for S/D (Additional file 1: Figures S3, S4, S8), suggesting that it is on the path to  $\text{Na}^+/\text{Na2}$  release as well.

These results for the large number of different constructs and conditions are remarkably consistent with the experimental evidence showing that the regular transport of the dopamine substrate (uptake) is affected differently by the various mutations/conditions than the reverse transport of this substrate (efflux) induced by AMPH. Thus, efflux is impaired by most of these mutations/conditions, with the exception of the S/D system, which exhibits dopamine efflux even in the absence of AMPH but under elevated intracellular  $\text{Na}^+$  concentrations [25]. In particular, experimental evidence points to the importance of  $\text{PIP}_2$ -containing membranes for various functional phenotypes of hDAT, including AMPH-induced efflux, but shows that substrate transport is only mildly affected if  $\text{PIP}_2$  content is reduced [34–36].

The dependence of functional properties of the eukaryotic transporters on  $\text{PIP}_2$  is not shared by the bacterial transporter homologs, such as the structural prototype LeuT [16, 37], which do not require  $\text{PIP}_2$ -containing membranes for transport and also do not exhibit reverse transport. We reasoned that comparing molecular details of functional mechanisms involving the N-terminus in the presence and absence of  $\text{PIP}_2$  would shed new light on the role introduced in evolution by the long N-terminus of the eukaryotic transporters. To discern the source of underlying mechanistic differences that connect  $\text{PIP}_2$  sensitivity to the long N-terminus, it therefore became necessary to understand (1) how the initiating step of substrate transport, i.e., the release of  $\text{Na}^+/\text{Na2}$ , is achieved in  $\text{PIP}_2$ -containing vs  $\text{PIP}_2$ -depleted membranes, and (2) what the role of the N-terminus interactions is in the CCRF (including  $\text{Na}^+/\text{Na2}$  release process) when  $\text{PIP}_2$  is not present. To this end we used the MSM analysis to obtain a kinetic model for the  $\text{Na}^+/\text{Na2}$  release process in the no- $\text{PIP}_2$  system, as the comparison of quantitative terms for the wild-type protein with/without  $\text{PIP}_2$  allows robust mechanistic inferences as illustrated below.

#### Markov state model analysis of $\text{Na}^+/\text{Na2}$ release in $\text{PIP}_2$ -depleted membranes

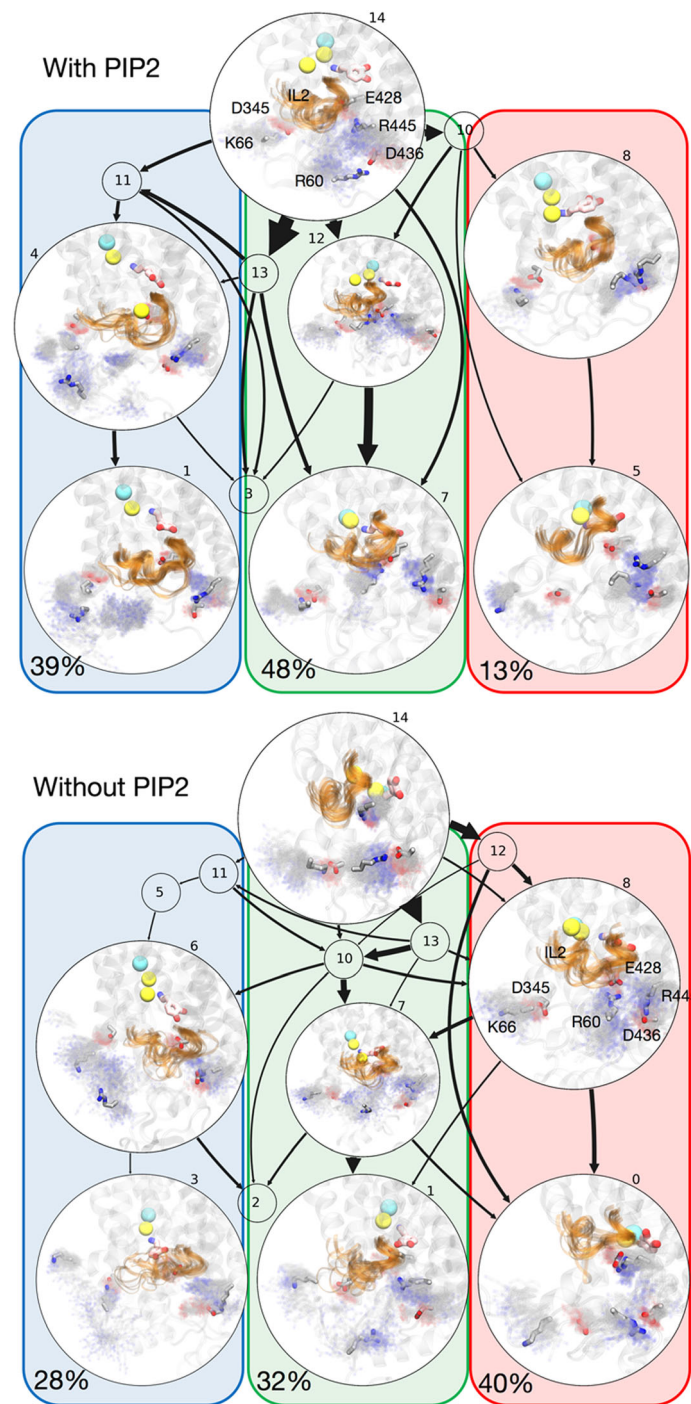
To enable direct comparison of the results with the MSM analysis of wild-type hDAT in  $\text{PIP}_2$ -containing membranes [32], we built and analyzed the MSM for the no- $\text{PIP}_2$  system following the same protocol (see [Methods](#) and Ref. [32]). Thus, as the same mechanism was followed in the two compared conditions (i.e., with/without  $\text{PIP}_2$ ), the same set of parameters as before [32] (Additional file 2: Table S3) was used to generate the reduced conformational space with the time-structure-based independent component analysis (tICA) method (see [Methods](#)). The tICA energy landscape (Fig. 3b) was obtained by projecting all the conformations from all the trajectories onto the first two tICA reaction coordinates. Visualization of conformations belonging to different regions of the tICA energy



landscape revealed that this landscape, unlike the one for PIP<sub>2</sub>-containing membrane conditions (shown in Fig. 3a), could be divided into only two (rather than three) regions in terms of the location of the Na<sup>+</sup>/Na<sub>2</sub> ion: one in which Na<sup>+</sup>/Na<sub>2</sub> is still bound in the Na<sub>2</sub> site, and the other in which Na<sup>+</sup>/Na<sub>2</sub> is already released (Fig. 3b). Thus, this tICA space (Fig. 3b) does not contain a region representing the intermediate state seen in the wild-type hDAT system in PIP<sub>2</sub>-enriched membranes when Na<sup>+</sup>/Na<sub>2</sub> has left the binding site but is not yet released to the intracellular environment because it is interacting with the E428 side chain (Fig. 3c). Because this interaction requires the E428 side chain to be free from its partner in the E428–R445 gate [32], the results suggest that a change in N-terminus interactions due to PIP<sub>2</sub> depletion directly affects this gate. Indeed, the finding summarized in Fig. 2 shows a major reduction in the interactions of the N-terminus with IL4 in the absence of PIP<sub>2</sub>, which is thus seen to result in a more

stable R445–E428 gate in the absence of PIP<sub>2</sub> (Fig. 3d, see also Additional file 1: Figure S9).

To build the MSM from the two-dimensional (2D) tICA landscape shown in Fig. 3b, we followed the same protocol as before [32] to obtain the *implied time-scales* plot (see Methods). As shown in Additional file 1: Figure S10 (*top panel*), Markovian behavior of the system is observed after a lag time of  $\sim 80$  ns; therefore, the transition probability matrix (TPM) (see Methods) at an 80-ns lag time was selected for all subsequent analyses of the no-PIP<sub>2</sub> system. Mapping all microstates on the tICA landscape and coloring them based on the first MSM relaxation mode (i.e., the second TPM eigenvector, shown in Additional file 1: Figure S11) reveals that Na<sup>+</sup>/Na<sub>2</sub> is still bound in microstates with positive sign (red circles in Additional file 1: Figure S11), whereas the microstates with negative sign (blue circles in Additional file 1: Figure S11) have released Na<sup>+</sup>/Na<sub>2</sub> to the intracellular environment.



**Fig. 4** Comparison of Na<sup>+</sup>/Na<sub>2</sub> release pathways calculated in PIP<sub>2</sub>-enriched and PIP<sub>2</sub>-depleted membranes. Results from the transition path theory (TPT) analysis for the release of Na<sup>+</sup>/Na<sub>2</sub> to the intracellular environment by the wild-type hDAT in PIP<sub>2</sub>-containing membrane (*top*) and in PIP<sub>2</sub>-depleted membrane (*bottom*) systems. *Arrow thickness* represents relative flux magnitude for each transition. The numbers above circles show corresponding macrostate numbers. A representative conformation is shown for selected macrostates, with IL2 highlighted in orange (cartoon representation) on the transparent structure of hDAT. Density representations of the residues forming intracellular gates are shown as fuzzy background gray colors and the most probable locations of these residues within the density representation are highlighted in licorice rendering and labeled for reference. Dopamine is shown in pink licorice and sodium and chloride ions in yellow and cyan spheres, respectively. Red, green, and blue boxes highlight the three major release pathways of Na<sup>+</sup>/Na<sub>2</sub>; numbers indicate the fraction of the total flux carried by each major pathway

Since the state population flows from positive to negative states, the first MSM relaxation mode is seen to capture the overall release kinetics of Na<sup>+</sup>/Na<sub>2</sub>. The implied time-scale equation (see [Methods](#)) shows that this relaxation mode is characterized by timescales of ~1.1 μs, comparable to the previously reported kinetics for hDAT in the PIP<sub>2</sub>-containing membranes (~800 ns) [32] (Additional file 1: Figure S10).

To compare the mechanisms of sodium release from the Na<sub>2</sub> site of the wild-type hDAT in PIP<sub>2</sub>-containing vs PIP<sub>2</sub>-depleted membranes, we used the same transition path theory (TPT) analysis (see [Methods](#)) to obtain the most probable release pathways of Na<sup>+</sup>/Na<sub>2</sub>, and quantified the flux associated with each of these on a macrostate-based MSM using 15 macrostates as before [32]. Similar to the wild type in PIP<sub>2</sub>-containing membranes, several pathways are revealed in the no-PIP<sub>2</sub> system. Here, the first 10 pathways identified by the TPT analysis contribute ~80% of the total flux between Na<sup>+</sup>/Na<sub>2</sub> bound states and Na<sup>+</sup>/Na<sub>2</sub> released states (highlighted in Fig. 4; see Additional file 2: Table S5 for quantification of fluxes). Their structural context is shown in Fig. 4 and Additional file 1: Figure S13.

The identification of the major Na<sup>+</sup>/Na<sub>2</sub> release pathway in the no-PIP<sub>2</sub> system reveals a clear difference from the release in the PIP<sub>2</sub>-enriched system (cf. panels labeled “with PIP<sub>2</sub>” and “without PIP<sub>2</sub>” in Fig. 4) that echoes the difference in the modes of interaction described by the results in Fig. 2. In the no-PIP<sub>2</sub> system, the first major pathway for the release contributes ~40% of the total flux and is formed when both the R60–D436 and the E428–R445 gates remain closed (a consequence of the much lower interaction with IL4 as seen in Fig. 2), but the K66–D345 is open (highlighted in the transparent red box in Fig. 4). This is in sharp contrast to the result in the presence of PIP<sub>2</sub> where this pathway is a minor contributor to the total flux, at only ~13%.

The second major pathway in the no-PIP<sub>2</sub> system is enabled when both the R60–D436 and E428–R445 gates are broken, but R60 is now able to form interactions with E428 (Fig. 4, green box). This pathway contributes ~32% to the total flux. Notably, this was the major Na<sup>+</sup>/Na<sub>2</sub> release pathway in the presence of PIP<sub>2</sub>, contributing about 48% percent of the total flux. But because in the absence of PIP<sub>2</sub> R60 (or R445) is interacting with E428, the intermediate state cannot form.

The third major pathway in the no-PIP<sub>2</sub> system is formed when the R60–D436 gate is broken and the N-terminus moves closer to IL2. The intracellular gate interaction E428–R445 is still maintained (Fig. 4, blue box). In the presence of PIP<sub>2</sub>, this pathway contributes about ~39% to the total flux, but in the no-PIP<sub>2</sub> system the contribution is reduced to 28%. Importantly, a major difference is that in the presence of PIP<sub>2</sub>, R60 is seen to be engaged in PIP<sub>2</sub>-

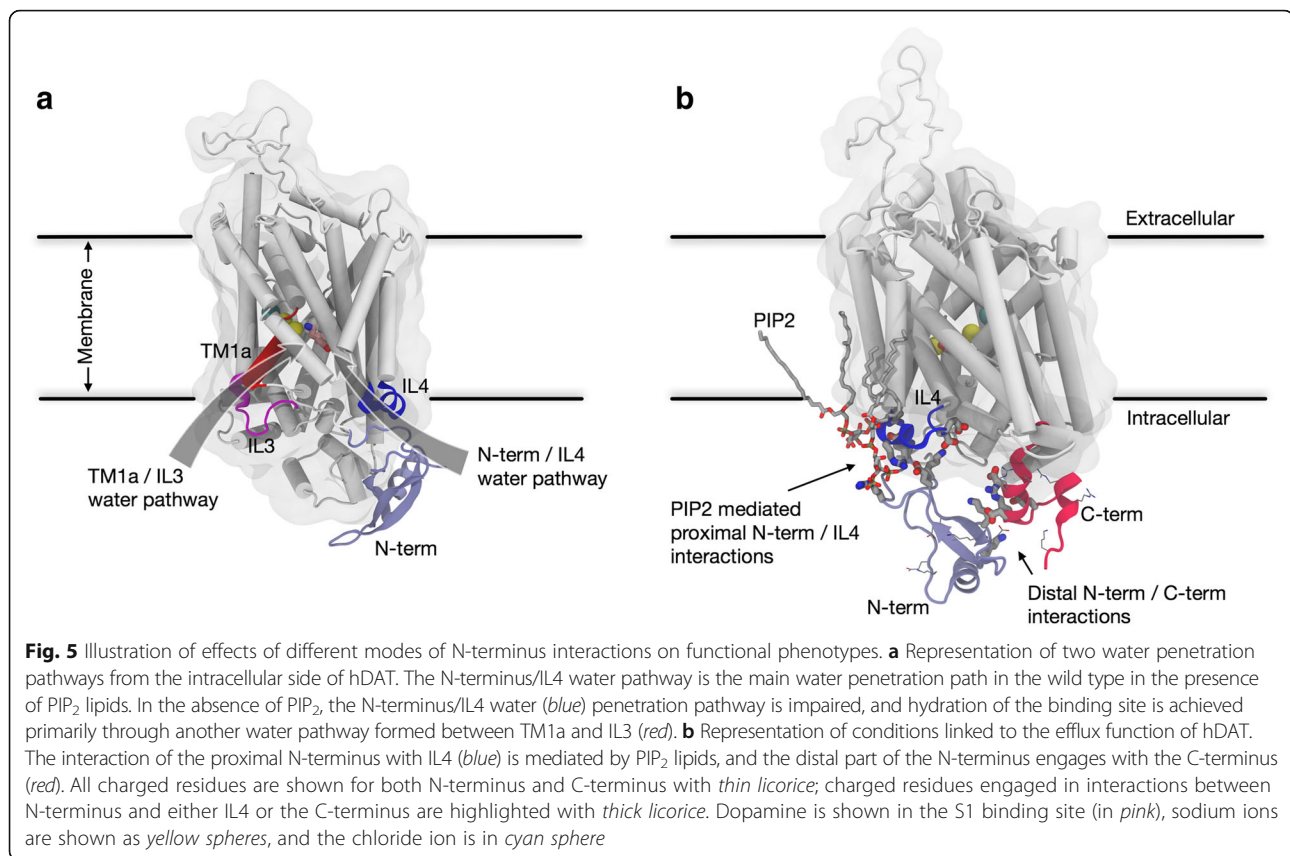
mediated interactions with several positively charged residues from IL2 (K257, K260, K264), whereas in the no-PIP<sub>2</sub> system the association between the N-terminus and the IL2 region still permits interactions between R60 and D68 and/or D345 (see macrostate 3 in Fig. 4 and Additional file 1: Figure S13).

Overall, the TPT analysis shows that the preference ranking of the various release pathways, which we quantified by calculating fluxes, is strongly affected by the PIP<sub>2</sub> lipids in a manner consistent with the modes of interaction with the N-terminus. The difference in Na<sup>+</sup>/Na<sub>2</sub> mechanism between the two conditions (with/without PIP<sub>2</sub>) is underscored by the finding that the increased dynamics of the N-terminus (Additional file 1: Figure S12) in the absence of PIP<sub>2</sub> results in a destabilization of the K66–D345 gate, which enhances the Na<sup>+</sup>/Na<sub>2</sub> release flux through a pathway that was only marginally active in the presence of PIP<sub>2</sub> (pathway highlighted in red box). Taken together, these results show how the eukaryotic transporters can adapt to different membrane composition conditions by utilizing different N-terminus interaction patterns so that release of Na<sup>+</sup> from the Na<sub>2</sub> site is maintained.

## Discussion

The extensive investigations of the molecular mechanisms underlying the vital role of hDAT in signal transduction have profited much from the availability of structurally simpler prototypes of the NSS family, such as the bacterial analog LeuT (a leucine and alanine transporter), for which the crystallographic data provided the first structural basis for detailed molecular studies (see [16, 37]). When the striking fold similarity of LeuT to the eukaryotic and human neurotransmitter transporters such as DAT was established [38, 39], it enabled major progress in understanding the functional mechanism, as documented in a very large number of publications (for reviews see [40]). It became clear, however, that various physiologically important mechanisms that eukaryotic transporters such as DAT and SERT have acquired through evolution involve allosteric coupling to their environment that differentiate them from the structurally simpler bacterial analogs. In a large number of studies [20, 22, 24, 25, 29, 32, 34, 41–50] to which we have contributed findings from both computation and experiments, these new functions of the eukaryotic transporters — such as regulation by lipids and by phosphorylation, and the observed efflux phenotype — were shown to be mechanistically modulated by defined structural elements, especially the relatively long N- and C-terminal segments that are found exclusively in this class, and not in bacterial counterparts [39]. These intracellularly located N- and C-terminal segments are the most divergent structural units within the NSS family of proteins, as they range in size from just a few (~10) amino acids (in bacterial members, such as LeuT) to segments





containing > 60 residues (N-terminus of eukaryotic NSS such as DAT, SERT; see [39, 51]).

To understand how the new functions of the eukaryotic transporters relate to the presence of the N-terminus and its dynamic properties, we quantified the interaction of the hDAT N-terminus with the intracellular regions of the hDAT transporter. On this basis we were able to relate different modes of interaction to the functional mechanisms of DAT through the relation between the experimentally measured effects of mutations/conditions and the changes in modes of interaction. The release of the Na<sup>+</sup> ion from the Na2 site, which is known to be a key first step in the substrate translocation cycle [26, 31], served as an established functional readout for the initiation of the transport cycle. We showed how the effects of modes of interaction of the N-terminus with various intracellular regions of the transporter relate to various modes of intracellular gate opening and paths of water penetration. This led to a mechanistic interpretation of the experimentally measured modifications of the functional properties observed for mutant constructs in the presence and absence of PIP<sub>2</sub>. Together, our results reveal how changes in the modes of interaction stemming from N-terminus mutations and PIP<sub>2</sub> depletion are directly associated with stabilization/destabilization of intracellular gates, and also show their

effect on the penetration of water into the binding site (Fig. 5a) that is required for uptake and efflux in hDAT.

By specifying the modes of interactions of the N-terminus that regulate the opening probabilities of water pathways separately for the *distal* and the *proximal* segments of the N-terminus, our findings provide a clear mechanistic explanation for a persistent conundrum in the literature regarding the effects of (1) truncating the first 22 residues of the N-terminus, and (2) PIP<sub>2</sub> depletion, and in particular their different effects on the uptake and efflux aspects of eukaryotic transporter function. That the truncation has little effect on uptake is explained by our finding (summarized in Fig. 2) that the pattern of interactions of the N-terminus with IL4 of DAT is mimicked by just the *proximal* segment, without special involvement of the *distal* segment. However, this interaction, which has been shown to disrupt the E428–R445 gate and thus serves as one of the major mechanistic triggers for Na<sup>+</sup>/Na<sub>2</sub> release, is PIP<sub>2</sub>-mediated [29] and is reduced under no-PIP<sub>2</sub> conditions (Fig. 2). This might have suggested a reduced release of Na<sup>+</sup>/Na<sub>2</sub> that would impair the substrate uptake function under these conditions. Yet this is not what is observed experimentally, as the uptake function of hDAT is not impaired by PIP<sub>2</sub> depletion from the membrane. The mechanistic explanation emerges from our

analysis of (1) the simulations presented here that show how modes of interaction of the N-terminus change under the different conditions (and for the various mutant constructs; Fig. 2), and (2) the MSM from the simulation trajectories of the no-PIP<sub>2</sub> condition showing how Na<sup>+</sup>/Na<sup>2</sup> release is maintained, but with different probabilities of the release pathways that involve alternative water penetration channels (e.g., the TM1a/IL3 channel (Fig. 5a) that is formed when K66–D345 interaction breaks).

The effect of N-terminus truncation on efflux is even more interesting in the context of the functional difference between the bacterial and eukaryotic transporters, because elimination of the *distal* segment in the neurotransmitter SLC6 transporters impairs a function not shared with the bacterial homologs. The relation of the *distal* segment to efflux makes it tempting to speculate that some combination of the interaction modes of the *distal* and *proximal* N-terminus (Fig. 5b) is required for efflux. Indeed, the results summarized in Fig. 2 show (1) a reduction of *proximal* N-terminus/IL4 interactions in constructs with low efflux activity (R51W and K3/5A) and in PIP<sub>2</sub>-depleted membranes and (2) a reduction in *distal* N-terminus/C-terminus interactions in the efflux-deficient K3/5A mutant, and an increase of these interactions in the efflux-promoting S/D construct. An involvement of the *distal* N-terminus/C-terminus interactions in facilitating efflux is also consistent with the observation that palmitoylation of the C-terminus reduces efflux, presumably by limiting interaction with the C-terminus. This limitation would reduce N-terminus phosphorylation [52, 53] by the Ca<sup>2+</sup>/calmodulin-dependent protein kinase II (CaMKII) that is proposed to attach to the distal C-terminus of hDAT in order to phosphorylate the serine residues in the distal N-terminus for AMPH-induced efflux [25].

## Conclusions

The molecular mechanisms emerging from the studies presented here offer detailed quantitative explanations for the experimental findings about the modulation of transporter function by the N-terminus of DAT, SERT, and other eukaryotic transporters. These molecular mechanisms connect the observed elongation of N- and C-termini in the evolution from bacterial homologs, with the appearance of new functional properties (e.g., efflux) that are not shared by the bacterial homologs which lack the long N-terminus but share the overall molecular architecture (termed the “LeuT-fold” [37]). They are clearly enabled by specific roles of either the *proximal* or *distal* segments, or both working in concert. Notably, the specific (different) involvements of *proximal* and *distal* segments — such as the role of the *proximal* segment in sustaining transport in PIP<sub>2</sub>-depleted membranes and that of the *distal* segment in modulating efflux — may represent an evolutionary adaptation required for the function of eukaryotic transporters expressed in

various cell types of the same organism, which can differ in the lipid composition and protein complement of their membrane environment.

## Methods

### System preparation

The molecular model of full-length wild-type hDAT used in this study is the same as that described and investigated earlier [32]. The R51W and K3A + K5A constructs were prepared by introducing the mutations in the wild-type hDAT model using the VMD mutator plugin [54]. To build the S/D mutant (simultaneous mutations of S2, S4, S7, S12, and S13 residues to Asp), we combined, using Modeller version 9v1 [55], the 57–620 residue stretch from the wild-type hDAT structure with the structural model of the 1–57 S/D segment elaborated and described previously [20].

The full-length models of the hDAT constructs R51W, K3A + K5A (heretofore referred to as “K3/5A”), and S/D were inserted into the same pre-equilibrated compositionally asymmetric bilayer membrane used for MD simulations of the wild-type hDAT [32]. This lipid bilayer was designed to mimic a neuronal cell plasma membrane and contains 5% PIP<sub>2</sub> lipid on the intracellular leaflet of the bilayer (see Additional file 2: Table S1 for the membrane lipid composition). For the simulations of the wild-type hDAT in the PIP<sub>2</sub>-depleted membrane environment, as done previously [29], all the PIP<sub>2</sub> lipids in the bilayer were changed to 1-palmitoyl-2-oleoyl-*sn*-glycero-3-phosphoethanolamine (POPE) lipids, the major component of the intracellular leaflet of our model bilayer. All the hDAT-membrane systems were solvated in a 150 mM K<sup>+</sup>Cl<sup>-</sup> TIP3P water solution with ions added for neutrality, resulting in a final atom count of ~ 150,000.

### Molecular dynamics simulations

All-atom MD simulations were carried out using the same scheme as described earlier for the wild-type hDAT in PIP<sub>2</sub>-enriched membranes [32]. Briefly, using NAMD software version 2.10 [56], the systems first were equilibrated following the same multi-step equilibration protocol used previously [32] during which the backbone of the protein was first fixed, then harmonically restrained, and finally released. After this equilibration phase, the velocities of all the atoms in the system were reset (at  $T = 310$  K using random number seed), and 50 independent ~ 1- $\mu$ s-long unbiased MD simulations were carried out using the latest version of the ACEMD software [57], resulting in a cumulative MD simulation time of ~ 50  $\mu$ s per system. These production simulations were performed under NVT ensemble and with all the default run parameters validated by the ACEMD developers (<https://www.acelera.com/>) and in a large number of published

applications (e.g., see <https://www.acellera.com>). The run parameters (4-fs time step with hydrogen mass repartitioning; particle mesh Ewald (PME) for electrostatics; switched Lennard-Jones interactions with a cutoff of 9 Å, and switching distance set to 7.5 Å) have been shown to reliably reproduce known values for free energy of protein folding and a variety of properties of lipid membranes [58, 59]. In addition, ensemble MD simulations with ACEMD have been generally used to generate large data sets of trajectories for quantitative analysis of kinetics of ligand-induced conformational transitions in G protein-coupled receptors (GPCRs) [60], of protein-protein association/dissociation processes [61], of phospholipid scrambling processes mediated by the GPCR opsin [62], as well as for identifying pathways for spontaneous cholesterol movement in adenosine A2A GPCR [63].

### Calculation of the interaction strengths

To obtain a measure of interaction between the N-terminus and other intracellular regions of hDAT, we counted the number of interactions between charged residues from the N-terminus and the intracellular loop regions. The number of interactions was quantified for a cutoff distance of 7 Å between interacting residue pairs from head group atoms of the N-terminus and of intracellular domain residues (using  $N_{\xi}$  for Lys,  $C_{\xi}$  for Arg,  $C_{\delta}$  for Glu, and  $C_{\gamma}$  for Asp). The residues included in the calculations are listed in Additional file 2: Table S2. All 50 trajectories for each construct were used for these calculations, with the first 500 ns of each trajectory considered an equilibration phase so that only the 500–940 ns time interval from each trajectory was used. This ensured that the total number of frames used for the analysis was the same for each construct. The total number of interactions was then divided by the total number of trajectories (i.e., 50) to obtain the average “interaction strength” represented in Fig. 2. Error bars were calculated by taking the standard deviation of “interaction strength” in all 50 trajectories.

### Markov state-based quantitative kinetic model construction

We used the Markov state model (MSM) approach to analyze the trajectories in the absence of PIP<sub>2</sub> lipids and build quantitative kinetic models of sodium release from the Na2 site for comparison with the kinetics and pathways calculated in the presence of PIP<sub>2</sub> in Ref. [32]. Therefore, we have followed the same protocol here as described in detail in Ref. [32]. Such quantitative kinetic models provided by MSMs [64–66] have been widely applied in protein folding studies [67, 68], and MSM-based kinetic model predictions have been validated experimentally [69, 70]. We and others have used MSMs combined with reactive flux

analysis, such as transition path theory (TPT) analysis, to obtain key mechanistic insights into functions of membrane proteins [32, 71, 72]. The three main components for achieving quantitative MSM-based kinetic models are briefly reviewed below.

1. Dimensionality reduction using tICA. Reducing the dimensionality of a system as large and complex as the membrane-immersed hDAT is necessary in order to construct an interpretable kinetic model. A natural choice of suitable reaction coordinates is those that can project the conformational space of the system along its *slowest* reaction coordinate, as this reaction coordinate will capture most of the conformational heterogeneity during the time course of the simulation [73]. The *time-structure-based independent component analysis* (tICA) method was developed recently for this purpose of capturing the slowest reaction coordinate of a system [74–76]. Briefly, the tICA method involves a transformation that utilizes two matrices constructed from the trajectory data: the covariance matrix  $\mathbf{C}$ , and a time-lagged covariance matrix  $\mathbf{C}_{TL}$ . The slowest reaction coordinates of a system are then defined by eigenvectors of the generalized eigenvalue problem  $\mathbf{C}_{TL}\mathbf{V} = \mathbf{C}\mathbf{V}\mathbf{\Lambda}$ , where  $\mathbf{\Lambda}$  and  $\mathbf{V}$  are the eigenvalue and eigenvector matrices, respectively. The eigenvectors corresponding to the largest eigenvalues identify the slowest reaction coordinates. Here we used a lag time of 16 ns to construct the time-lagged covariance matrix  $\mathbf{C}_{TL}$ , and the tICA parameters were chosen as before for the hDAT molecular system [32] to measure (a) the dynamics of the Na<sup>+</sup> ion from the Na2 site, termed Na<sup>+</sup>/Na2, and (b) the dynamics of intracellular gates formed between residues R60, D436, R445, and E428 (Additional file 2: Table S3).
2. Markov model construction. The conformational transitions of biomolecular systems where the time between transitions is long enough can be modeled as Markov chains [64] in which transitions among states depend only on the current state of the system (i.e., Markovian behavior). Such Markov models provide powerful tools for outcome prediction by enabling the extraction of long time-scale information from multiples of short time-scale events. Two components needed for the construction of such a Markov model are an ensemble of *microstates* of the system, and of the transitions among these microstates [77]. Microstates are defined by clustering the conformational space of the system into several basins using automated clustering algorithms like *K*-means or *K*-centers, and this technique is most practical if performed in a dimensionality-reduced space such as the one

obtained from the tICA transformation. The transitions among the microstates are calculated for a particular time interval between each of the transitions (called the lag time) and stored in the *transition count matrix*. By row-normalizing the transition count matrix one obtains the *transition probability matrix* (TPM). To validate Markovian behavior, the TPMs are constructed for multiple lag times and the relaxation timescales of the system are extracted by using the relation:

$$\tau_i = -\frac{\tau'}{\ln \lambda_i}$$

where  $\tau'$  is the lag time used for building the TPM,  $\lambda_i$  is the  $i^{\text{th}}$  eigenvalue of the transition probability matrix, and the resulting  $\tau_i$  is called the *implied timescale* corresponding to the  $i^{\text{th}}$  relaxation mode of the system. The system is considered to be Markovian if the behavior is such that  $\tau_i$  is independent of  $\tau'$ ; the corresponding TPM is a *Markovian* TPM that contains all the information about the thermodynamics and kinetics of the system. Thermodynamic information is stored in the first eigenvector (which has an eigenvalue of 1). Information about kinetics is stored in the subsequent (second, third, etc.) eigenvectors, with the second eigenvector (corresponding to the second largest eigenvalue) capturing the slowest dynamics of the system, the third eigenvector capturing the second slowest dynamics, and so on. Following the same protocols as described in detail in Ref. [32] for the construction of the Markov models, we discretized the reduced conformational space generated by the first two tICA reaction coordinates into 100 microstates (Additional file 1: Figure S11) using the *K-means* clustering algorithm implemented in the MSMBuilder3 software [78]. TPMs were constructed at several different lag times to obtain the implied time-scale plots shown in Additional file 1: Figure S10, so that the one in which Markovian behavior is exhibited can be identified and chosen for further analysis.

3. Transition path theory analysis. In addition to the thermodynamics and kinetics information it contains, the Markovian TPM also contains mechanistic information for a dynamic system. An established means of revealing such mechanistic information inherent in the TPM is the TPT analysis that identifies the most probable flux pathways of the system [79]. TPT provides such pathways by constructing a *flux matrix* from the Markovian TPM. This matrix conversion has been documented in detail [79, 80], and its implementation is discussed in our previous

publication [32]. Although directly applicable to MSM in the microstate space (on the order of hundreds to thousands of states), TPT analysis is usually done on a *macrostate* MSM (on the order of tens of states) for a better visualization of flux pathways. Here we transitioned from the microstate MSM to the macrostate MSM by using the robust Perron cluster analysis (PCCA<sup>+</sup>) algorithm [81] that lumps microstates into macrostates using the sign structure of the MSM eigenvectors (this assumes that microstates with the same signs, positive or negative, in the MSM eigenvectors, will have similar kinetics [82]). Using the PCCA<sup>+</sup> algorithm we obtained 15 macrostates, and by applying the TPT analysis to these macrostates we obtained the most probable flux pathways for the system.

## Additional files

**Additional file 1:** Supplementary Figures. **Figure S1.** Na2 distance time evolution for no-PIP2 system. **Figure S2.** Na2 distance time evolution for R51W system. **Figure S3.** Na2 distance time evolution for S/D system. **Figure S4.** Na2 distance time evolution for K3A/K5A system. **Figure S5.** Intracellular gating residues distances for S/D system. **Figure S6.** Intracellular gating residues distances for R51W system. **Figure S7.** Intracellular gating residues distances for no-PIP2 system. **Figure S8.** Frequencies of intracellular gating residues interactions. **Figure S9.** Distribution of the R445–E428 distance in different constructs of hDAT. **Figure S10.** MSM implied-timescales plots. **Figure S11.** MSM 1st relaxation mode for no-PIP2 system. **Figure S12.** Localization density of the N-terminus near the membrane. **Figure S13.** MSM macrostates for the no-PIP2 systems. (PDF 9.96 mb)

**Additional file 2:** Supplementary Tables. **Table S1.** Lipid composition of the model membrane. **Table S2.** Residues included in the calculation of interaction strength between N-terminus and intracellular regions. **Table S3.** Parameters used for dimensionality reduction and tICA construction. **Table S4.** Correlation values for water coordination number. **Table S5.** MSM predicted sodium release pathways. (PDF 169 kb)

## Abbreviations

DAT: Dopamine transporter; hDAT: Human dopamine transporter; MSM: Markov state model; NSS: Neurotransmittersodium symporter; SERT: Serotonin transporter; SLC6: Solute carrier 6; tICA: Time-structure-based independent component analysis; TM: Transmembrane; TPM: Transition probability matrix; TPT: Transition path theory

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## Availability of data and materials

All simulation trajectories are available upon request.

**Authors' contributions**

All authors contributed to conceiving the project and experimental design, AMR did the analysis and carried out the MSM construction, GK carried out the MD simulations, and all authors participated in the interpretation of results and in writing the manuscript. All authors read and approved the final manuscript.

**Ethics approval and consent to participate**

Not applicable.

**Consent for publication**

Not applicable.

**Competing interests**

The authors declare that they have no competing interests.

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